

## Ammotragus lervia. By Gary G. Gray and C. David Simpson

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### *Ammotragus* Blyth, 1840

*Ammotragus* Blyth, 1840:13. Proposed as a subgenus of *Ovis*.  
Type species *Ovis (Ammotragus) tragelaphus*.

*Aegoceros* Heuglin, 1861:16. Type species *Aegoceros tragelaphus*.

**CONTEXT AND CONTENT.** Order Artiodactyla, Suborder Ruminantia, Infraorder Pecora, Family Bovidae, Subfamily Caprinae, Tribe Caprini. The genus *Ammotragus* consists of one species, as treated below.

### *Ammotragus lervia* Pallas, 1777

Barbary Sheep; Aoudad, Audad, Udad, Arui;  
Mouflon à Manchettes; Mähnenspringer;  
Muflone Berbere; Afrui

*Antelope lervia* Pallas, 1777:12. Type locality "Africæ borealiori propria."

*Ovis tragelaphus* Afzelius, 1815:216. Type locality "Mauritanie."

*Ovis ornata* I. Geoffroy Saint-Hilaire, 1827:264. Type locality "près des portes de la ville du Caire" (=Caire, Egypt).

*Ammotragus lervia* Thomas, 1902:13, first use of current name combination.

**CONTEXT AND CONTENT.** Context noted in generic summary above. Six subspecies are recognized (Allen, 1939) as follows:

*A. l. lervia* (Pallas, 1777:12). Type locality restricted to the "Department of Oran, western Algeria" by Harper (1940:327).

*A. l. ornata* (I. Geoffroy Saint-Hilaire, 1827:264), see above.

*A. l. sahariensis* (Rothschild, 1913:459). Type locality "Oued Mya" between El-Golea and In-Salah, 28°30'N, 3°E, Algerian Sahara.

*A. l. blainei* (Rothschild, 1913:460). Type locality "Border of Dongola Province and Kordofan," Anglo-Egyptian Sudan.

*A. l. angusi* (Rothschild, 1921:75). Type locality "Tarrouaji Mt., Asben, 3,100 ft.," French West Africa.

*A. l. fassini* (Lepri, 1930:271). Type locality Garian range, northwestern Libya.

**DIAGNOSIS.** The following combination of gross morphological characteristics is diagnostic for *Ammotragus*: presence of true horns, subcaudal gland, ventral neck mane, and chap hair on front legs of adults (particularly evident on males); absence of preorbital, interdigital, or inguinal glands, and inter-ramal chin beard.

**GENERAL CHARACTERS.** Pelage is rufous or tawny brown with occasional dark brown areas about the head and forequarters. The chin, belly, and inside of legs are whitish, and a circular spot of white hairs frequently occurs on the head between the horns. A fringe of long hair, the mane, extends from under the throat down the front of the neck to the brisket. This hair then bifurcates and continues down the forelegs of mature animals, where it is termed chaps. Although a mane is found on other wild sheep, chaps are peculiar to aoudads. A short erect fringe is present on the back from the base of the neck to just behind the withers.

The head is relatively long with narrow, evenly tapered ears and a small black nose. Eyes are large and distinctive; a horizontal pupil is surrounded by a yellow iris.

True horns (Fig. 1), present on both sexes, are relatively large and moderately long, with a moderately high spiral angle (small radius). They are elliptical and keeled in cross section, with a fairly broad frontal surface, exhibit heteronym winding, and have numerous shallow and uniform sulci (rings) as well as periodic growth checks or annuli (Ogren, 1962, 1965; Geist, 1966; Schaffer and Reed, 1972). Horn points are directed inward, or inward and downward. Horn lengths and basal circumferences

rarely exceed 840 and 355 mm in males, and 510 and 205 mm in females.

The neck is robust and the forequarters are more massive than the hind, particularly in males. This appearance is accentuated by the topline which slopes from the withers. The legs appear to be short and stocky, and hooves are large, blunt, and well developed. Dew claws are usually hidden by chap hair. The tail is 175 to 205 mm long, tufted on the terminal half, and naked underneath (McClellan, 1955). Preorbital, interdigital, and flank glands are absent.

Dimensions of the largest males harvested in New Mexico (Ogren, 1965) were: total length, 196 cm; height, 112 cm; girth, 137 cm; weight, 145 kg. The largest females measured 165 cm, 94 cm, 112 cm, and 63.5 kg. These are significantly larger measurements than have been reported for type specimens from Africa, which were from 94 to 99 cm in height (Clark, 1964).

**DISTRIBUTION.** Lhote (1957:88) designated the range as being all the hills of the Sahara, but particularly the Hoggar, Tassili, Ayr, and Tibesti mountains. Barrett (1967) summarized the endemic geographic distribution, indicating that the species inhabited all the major mountain massifs of North Africa above 15° latitude (Fig. 2), citing Bigourdan and Prunier (1937), Rode (1943), Brouin (1950), Malbrant (1952), and Edmond-Blanc (1957). Joleaud (1927) also delineated the distribution.

Barbary sheep were present in some European zoological gardens during the last half of the previous century (Ogren 1965:5), were introduced into the Teutoburger Forest of West Germany (Strasson, 1916), and onto Italian game preserves (Zammarano, 1930). The first importations from Europe to zoos in the United States occurred about 1900 (Ogren, 1965:6). Barbary sheep readily reproduce in confinement, and surplus zoo stock was eventually sold to private parties. Wild populations in the southwestern United States were derived from groups which were released or escaped from private herds. The larger wild populations are located in Palo Duro Canyon of the Texas Panhandle, the Canadian River Gorge, Canyon Largo, and Hondo Valley of New Mexico, and Santa Lucia mountains of west-central California.

**FOSSIL RECORD.** Fossil sheep remains recovered from superficial deposits in Europe were referred to as *Ovis tragelaphus fossilis* by Serres (1848:149), and *O. magna* and *O. primoeva* by Gervais (1852:76). Lydekker (1912:310) commented that these "sheep were akin to the modern arui of North Africa." Osborn (1910:433), writing of the Pleistocene in North America, mentioned a "wild sheep (*Ovis paleotragus*) very similar to the existing Barbary sheep."

McBurney (1967) found remains of *Ammotragus* in Haua Feah, a large natural cave on the northern coast of Cyrenaican Libya, in a number of deposits which ranged in antiquity from about 85,000 to 2000 years B.P. Bate (1955) discussed the vertebrate faunas, including Barbary sheep, from Quaternary deposits in Cyrenaica. Vaufrey (1955) noted the species among the Pleistocene fauna in the Maghreb (Atlas massif), and Arambourg et al. (1934) recorded the taxon at Beni Segoual.

**FORM.** The skin is relatively thick, cutaneous glands are rare, and special dermal glands absent (Bourdelle, 1924). Females possess one pair of inguinal mammae.

In general, osteological characteristics are more nearly like those of goats than sheep (Bourdelle, 1924). Sexual dimorphism in horn size, sinuses, and associated cranial morphology is pronounced. Horn bases are displaced behind the orbit. Schaffer and Reed (1972) made the following observations on the cranial morphology of *Ammotragus*. The male skull is considered to be of an advanced shape among Caprini—that is, the foramen magnum is located beneath the horn bases and the back of the skull is more nearly vertical, so that the inclusive angle formed by the intersection of a plane along the parietal and supraoccipital bones with the plane of the palate is greater than 70°, and the ratio of

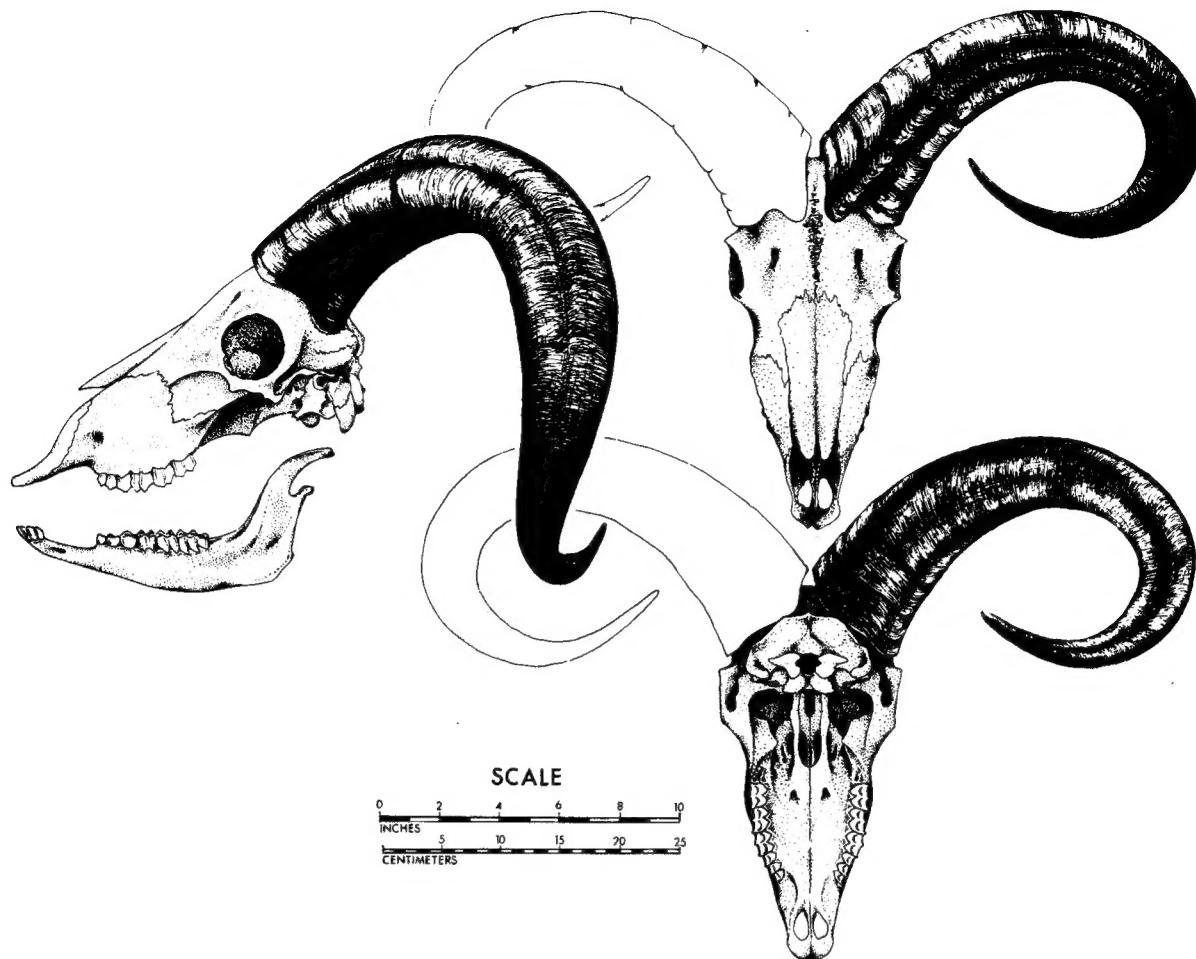


FIGURE 1. Dorsal, ventral, and lateral views of skull of male *Ammotragus lervia* from Palo Duro Canyon, Texas, U.S.A., owned by James Standridge. Drawings by C. David Simpson.

parietal length to ventral skull length approximates 0.15. In the aoudad this configuration has been achieved by a ventrad rotation of the brain case (Fig. 1). The frontal sinuses are extensive and the septa complex in males, moderately extensive and moderately complex in females; the cornual sinuses are extensive and septa complex in males, extensive and moderately complex in females. The vertebral formula is C 7, T 13, L 6, S 4, Cd 14 (Lydekker, 1913). Incisors are typically goat-like, whereas molars are more nearly sheep-like (Bourdelle, 1924). The dental formula is i 0/3, c 0/1, p 3/3, m 3/3, total 32.

The structure of the hemoglobin (Hb)  $\alpha$  polypeptide chain from this species consists of two forms, and both  $\text{I}^\alpha$  and  $\text{II}^\alpha$  chains closely resemble those of the domestic goat (Huisman, 1974); the  $\beta^B$  chain is apparently unique among the Caprini. The Amm-B Hb chain of Barbary sheep was shown to be related to but not identical with Hb B of the domestic sheep, and the Barbary sheep Amm- $\beta^C$ -chain was similar to the sheep  $\beta^C$ -chain (Huisman et al., 1968). The Hb of Barbary sheep was compared with Hb-I and Hb-II from Dutch Texel sheep and found almost identical to Hb-I in electrophoretic mobility, but of intermediate solubility between Hb-I and Hb-II in a series of phosphate buffers (pH 6.5) of graded molarities (Huisman et al., 1958). Hb-II has a greater affinity for oxygen, originated—in domestic breeds—from sheep living at high altitudes, and may be of adaptive significance; Hb-I is characteristic of breeds living at sea level. In an examination of blood proteins by electrophoresis in acrylamide and starch gel, *Ammotragus* was found to have five distinctly sheep-like biochemical characteristics, seven characteristics in common with both sheep and goats, and 10 unique characteristics (Manwell and Baker, 1977). Barbary sheep serum cholinesterase may be more primitive than either sheep or goat cholinesterase since it is more active on butyrylthiocholine and  $\alpha$ -naphthyl acetate.

The uterus is bicornuate and the semideciduous placenta cotyledonary; an unusual type of corpora luteal scar that Ogren (1965:62) frequently encountered was called a corpus veriformis.

Ogren (1965:65) described sperm structure and Steklenov (1972) compared sperm morphology with other representatives of the Caprinae.

A number of general morphological features can be used to separate *Ovis* from *Capra*, and Payne (1968) indicated that *Ammotragus* shares six of the nine characteristics he inventoried with *Capra*. Linear body measurements were listed by Rode (1943), Brouin (1950), and Panouse (1957).

**FUNCTION.** Brady and Ullrey (1975) presented serum biochemical and hematological parameters of weaned juvenile Barbary sheep fed an exotic ruminant diet. Serum biochemical and hematological values of captive adults more closely resembled values for white-tailed deer, *Odocoileus virginianus*, than those of cattle or sheep, and activities of three serum enzymes assayed were also similar to those reported for white-tailed deer (Tumbleson et al., 1970). Meyer (1967) and Huisman et al. (1968) also listed hematological values for *Ammotragus*, and Huisman and Miller (1972) demonstrated that Hb-B is replaced by another hemoglobin type (Hb-C) during bloodloss anemia whereas a Hb-C-like variant, Hb-C<sup>(na)</sup>, present in certain nonanemic Barbary sheep, cannot be replaced when the animal is made severely anemic.

Average daily gain in weight of weaned juveniles was not significantly affected by dietary protein level and the only significant dietary effect on blood parameters was on plasma urea nitrogen (Brady and Ullrey, 1975). The dietary protein requirement for weaned animals did not exceed 13.1% under study conditions, and levels up to 19.6% did not appear to be harmful.

Nouvel et al. (1969) reported the effect of an intramuscular injection of 12 ml of Sédalande (=Halo-anisone), fluoro-methoxyphenyl-piperazine butyrophenone, on a male; and Hampy (1978) immobilized three males with M99 (etorphine) using M50-50 (diprenorphine) as the antagonist drug.

**ONTOGENY AND REPRODUCTION.** Sperm is abundant in the epididymis of older males year-round (Ogren, 1965:64). Most matings are accomplished during an autumn rut, from September through November, but some take place throughout the year. The gestation period is 22 to 23 weeks (Brown, 1936; Lobanov and Treus, 1971), and up to 84% of births occur from March through May (Flower, 1932; Zuckerman, 1953; Lobanov and Treus, 1971). Fourteen mature females collected in New Mexico during the late autumn of 1957 had a total of 17 embryos (1.2 per female); the ratio was 1.6 embryos per female for seven mature females taken in the late winter of 1959 (Ogren, 1965:60). Of 67 females shot during the first eight hunts in Palo Duro Canyon, Texas, 44 (65.6%) were pregnant with 57 fetuses (1.3 per female). One female had triplets, 14 had twins, and 29 contained single fetuses (DeArment, 1971). Fetal sex ratio was 19 males:21 females, with 17 unknowns.

Twins are produced once in every six or seven births under conditions of confinement (Zuckerman, 1953; Blunt, 1963). The sex ratio at parturition is unity and birth weights average 4.5 kg (Blunt, 1963). The neonate is precocial and able to negotiate moderately rugged terrain almost immediately after birth (Ogren, 1965:69). Concurrent lactation and gravidity indicates that lactation overlaps the estrous cycle in some females.

Age of puberty is variable. Ogren (1965:65) found sperm in an 11-month-old male, but not in a 15-month-old male, and a known-age female produced an offspring when only 13 months, 12 days of age (Blunt, 1963). On two known occasions a female has produced offspring twice in less than 7 months, and one had three single births within 13 months, 4 days (Blunt, 1963). Ogren (1965:59) regarded all females at least 19 months old as being sexually mature.

Dental maturity is apparently achieved rather late. Irruption of the permanent canine, an incisiform tooth, was observed at 65 and 68 months in two known-age animals (Ogren, 1965:31). Longevity has exceeded 20 years in zoos (Ogren, 1965:6), but longevity in the wild probably rarely exceeds 10 years.

Although Gray (1954:60) indicated that no live births resulting from matings of *Ammotragus* males to female domestic goats (*Capra hircus*) had been produced, a viable female from this mating combination was reported by Petzsch (1957) and this hybrid was successfully backcrossed to a Barbary sheep male. Brentjes (1968) suggested that this provides a possible interpretation for the presence of goat-like animals in ancient Egyptian pictographs, and that such hybrids may have been relatively common because of the wide geographic distribution of Barbary sheep.

**ECOLOGY.** This species inhabits arid and semiarid lands from sea level to at least 945 m in elevation within its endemic North African range (Rothschild, 1921:75) and canyons with gorges to 305 m deep at elevations up to 1829 m in the southwestern United States (Ogren, 1965:10). Habitat utilization is greatest in areas of precipitous topography (Evans, 1967; Hampy, 1978).

The major spring food plant in New Mexico was mountain mahogany, *Cercocarpus breviflorus* (54.3% rumen content by volume), followed by grasses (25%), forbs (13.8%), and other browse (5.6%). Summer foods included *C. breviflorus* (31.5%), wavyleaf oak, *Quercus undulata* (30%), grasses (26%), forbs (8.8%), and other browse (3%). The autumn diet featured *Q. undulata* (54.3%), grasses (29%), forbs (12.4%), and other browse (4.1%); whereas the principal winter food category was grass (86%) supplemented by small amounts of browse (10.9%) and forbs (2.4%) (Ogren, 1962, 1965). Mountain mahogany and wavyleaf oak accounted for half the food volume of 79 plants identified. An analysis of stomach contents from 109 animals collected in late autumn or early winter hunting seasons in Palo Duro Canyon, Texas, from 1963 through 1970 (DeArment, 1971) revealed the following woody plants to be important dietary components during this season: sand shinnery oak, *Quercus havardii* (contained in 66.9% of the stomachs); honey mesquite, *Prosopis glandulosa* (53.2%); yucca, *Yucca angustifolia* (24.9%); mountain mahogany, *Cercocarpus montanus* (33.9%); juniper, *Juniperus* sp. (24.8%); netleaf hackberry, *Celtis reticulata* (13.8%); and black willow, *Salix nigra* (13.8%). Principal forbs were western ragweed, *Ambrosia psilostachya* (13.8%) and buffalobur, *Solanum rostratum* (10.1%); and grasses, little bluestem, *Andropogon scoparius* (18.4%), silver bluestem, *A. saccharoides* (15.6%), and blue grama, *Bouteloua gracilis* (14.7%). Research in progress at Palo Duro Canyon, based primarily on fecal pellet analysis, suggests that woody browse is the most important forage class in the spring (53% relative frequency), summer (68%), and fall (48%), while browse (42%) and grass (43%) are equally important during winter

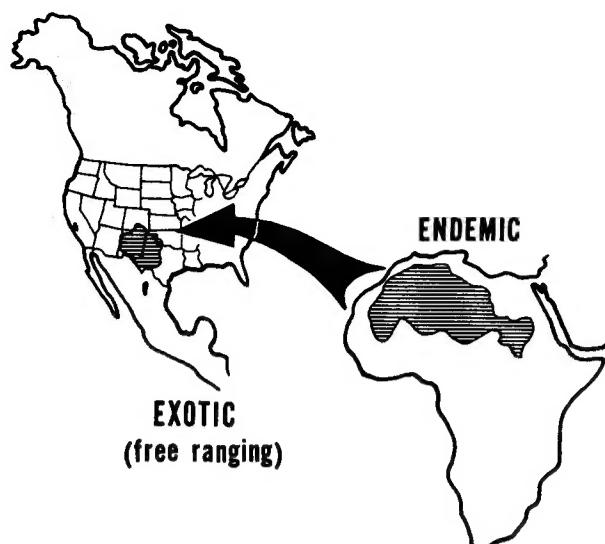


FIGURE 2. Endemic distribution of *Ammotragus lervia* in North Africa and exotic free-ranging distribution in North America. Map by C. David Simpson.

months (Simpson et al., 1978). Forbs were the preferred forage of a tame animal whose food habits were observed on the Edwards Plateau of Texas (Ramsey and Anderegg, 1972).

A few animals from an introduction at Palo Duro Canyon, Texas, dispersed up to 32.3 km (20 mi) from the release site within 5 to 7 months (DeArment, 1971). The home range of radio-collared adults in Palo Duro Canyon (Hampy, 1978) was 0.98 km<sup>2</sup> (0.38 mi<sup>2</sup>) and 2.64 km<sup>2</sup> (1.02 mi<sup>2</sup>) for one male during the winter as determined by minimum home range (MHR) and topographic home range (THR) methods, respectively. Summer MHR size was 19.26 km<sup>2</sup> (7.44 mi<sup>2</sup>) and summer THR was 30.85 km<sup>2</sup> (11.91 mi<sup>2</sup>). Two adult females exhibited a mean winter MHR of 2.09 km<sup>2</sup> (0.81 mi<sup>2</sup>) and THR of 4.57 km<sup>2</sup> (1.76 mi<sup>2</sup>); and these two plus one additional female had a mean summer MHR of 12.98 km<sup>2</sup> (5.01 mi<sup>2</sup>) and THR of 16.69 km<sup>2</sup> (6.44 mi<sup>2</sup>). Abrupt seasonal movements of as much as 23.4 airline km (14.5 mi) were also observed.

Interspecific competition has not been documented but it is likely that *Ammotragus* would have an advantage over most other ungulates in regions where such interactions are possible. Concern has been expressed for the future of the desert bighorn sheep (*Ovis canadensis*) in southwestern United States should Barbary sheep invade much of the desert bighorn's range (Barrett, 1967; Simpson et al., 1978). Predation does not appear to have a significant impact upon populations within its exotic range (Ogren, 1965:69; DeArment, 1971:15).

Thirteen helminth parasite species were reported from two Barbary sheep (Allen et al., 1956), and 17 helminth species were recovered from seven host animals (Allen, 1960), all collected in New Mexico. Only three helminth species were found in five hosts from Palo Duro Canyon, and worm burdens were light (Gray et al., 1978). Experimental cross-transmission of the abomasal worm, *Haemonchus*, suggested that the strain from wild Barbary sheep is less pathogenic than the strain from domestic sheep (Allen et al., 1958; Samson et al., 1964). A louse, *Bovicola neglecta*, described from zoo animals, is apparently host-specific for *Ammotragus* (Kéler, 1942:77; Werneck, 1950:81). The winter tick (*Dermacentor albipictus*) also was taken from New Mexico hosts (Allen et al., 1956; Allen, 1960).

Five of seven adult males from New Mexico showed positive reactors to the agar precipitin test for bluetongue (BT) (Trainer and Jochim, 1969); and serologic reactors to infectious bovine rhinotracheitis (IBR), BT, and epizootic hemorrhagic disease (EHD) were found in three, six, and three individuals, respectively, of 12 from the Palo Duro Canyon population (Hampy et al., 1979). Fifteen fatal cases of mycoplasmosis were recorded from Barbary sheep in the Frankfurt Zoo (Brack, 1966; Erns et al., 1972). Naturally occurring coronary and aortic atherosclerosis was discovered during necropsies of two adults which died at the St. Louis Zoo (Wallach and Middleton, 1970), and the lesions were more extensive than those reported to be naturally occurring in most other species. Seven animals died of Johne's disease, paratuberculosis (causative agent *Mycobacterium paratuberculosis*).



FIGURE 3. *Ammotragus* males fighting for dominance from a typical parallel position. Photo by D. Brent Hamby.

*tuberculosis*), also at the St. Louis Zoo, and two additional infected animals were euthanized (Boever, 1976).

Increasing numbers of people, improved transportation, the development of Saharan oil and mineral resources, and constant hunting pressure by some native tribes suggest that North African Barbary sheep populations levels will continue to decline. Indeed, the Egyptian subspecies, *A. l. ornata*, may already be extinct (Heinemann, 1972:493).

Nomadic peoples of the Sahara have depended on Barbary sheep for meat, hide, hair, sinews, and horns, and this species was considered to be of greater value to the economy of the Ahaggar Tuareg than other types of game (Nicolaisen, 1963:157). In the southwestern United States the species is important to the economy of game ranches, which produce animals for commercialized sport hunting (Schreiner, 1968; Teer, 1975). Wild populations in New Mexico and Texas provide challenging hunting opportunities rarely equaled by other big game, and supplement ranchers' incomes through the sale of hunting trespass rights, guide services, and use of camp facilities.

Traditional hunting and capture methods feature spears, stalking, disguises, artificial blinds or hides, use of dogs, running down with camels, and traps of the wheel or torsion type (Nicolaisen, 1963:158). In Texas, Barbary sheep have been roped with a lasso from a moving truck, driven into a net stretched across a game escape trail, or immobilized when struck by a drug-containing syringe dart fired from a Cap Chur gun out of a helicopter while in flight (Hamby, 1978). The most successful censuses have been accomplished using helicopters, although several population parameters can be estimated from repeated observation of groups at aggregation sites.

**BEHAVIOR.** Stable dominant-subordinate relationships existed between all members of the herd at the New York Zoological Park (Katz, 1949), and the social hierarchy was one of linear descent through males, females, and juveniles. The dominance status among juveniles seemed to be determined by the strength of the pair bond between juveniles and their mothers, as reflected by social distance between them. As the pair bond disappears, the dominance status of a juvenile establishes itself in its peer group. Group leadership was not related to dominance status, and the leader was always an adult female. Males were more timid when confronted with novel or potentially dangerous situations, and their following of females under such circumstances may partially reflect the conditioning of following females during conflict situations associated with the rutting season. A feed box experiment indicated differences in social distance between various pairs of males, reflected in the extent of food sharing and number of aggressive interactions.

The intraspecific threat posture consists of turning the head so the horns are directed toward a conspecific (Hamdy and

Schmidt, 1972). An alpha male also may intimidate conspecifics by raising the fringe hair over the withers, licking his muzzle, and uttering a low guttural growl. Intraspecific aggressive behavior is typically ritualized and of two major types (Katz, 1949; Haas, 1959; Schaffer and Reed, 1972:12). One, the head-on charge, is ordinarily confined to interactions between males and may feature a closing velocity up to 80.7 km/hr (50 mph) (Schaffer, 1968), with both animals remaining quadrupedal on impact. The other involves close butting and head shoving, or hooking the horns, neck, flank, or belly of the conspecific from a parallel position, followed by twisting or gouging motions (Fig. 3). The torque caused by such rotational movements has probably contributed to adaptive selection for musculature which resists rotation of the head (Schaffer, 1968), producing disproportionate development of the neck and forequarters in males. Fighting of both types was also common among females, but rarely involved charging an adversary. Displaced aggression was common, frequently continued chainwise to other individuals, and was influenced by physical proximity and lower social status of recipients (Katz, 1949).

Females defend against suitors during anestrus and are adept at striking them with their horns whether or not a male has mounted (Ogren, 1965:67). Proestrus females appear to excite males, which feed and ruminate less, sniff, or lick at female urine puddles and genitalia, and exhibit the typical bovid flehman (graphically pictured in Davis, 1974:262). Sexually aroused males may squat to urinate in female posture rather than the usual erect or forward-leaning stance, extrude and lick the penis, try to segregate a female by herding actions, and attempt mounting (Katz, 1949; Ogren, 1965:68). During estrus females sometimes lick the sides of a male, and mutual touching of muzzles is frequent. Copulation is accomplished by mounting, followed by several rapid thrusts, either immediately or several minutes after mounting, and is accompanied by male growls (Ogren, 1965:68). Copulation may be repeated up to 20 minutes later.

Juveniles exhibit a number of recognizable elements of aggressive and sexual behaviors, as well as typical maintenance and comfort postures (Haas, 1959), but these are frequently displayed in inappropriate contexts. Thus the ontogeny of behavior in this species may consist of rearranging many rather elaborate innate patterns into appropriate sequences, rather than a slow development or abrupt emergence of these patterns. Temporal periodicity in behavior embraces fairly regular cycles of alternating activity and rest within the daily regimen of herd behavior, as well as distinctive seasonal behavior (Katz, 1949; Haas, 1959).

**GENETICS.** This species has a karyotype of  $2N = 58$ ,  $FN = 60$ , composed of one pair of biarmed and 27 pairs of acrocentric autosomes, a large acrocentric X chromosome and a minute biarmed Y chromosome (Nadler et al., 1974). This karyotype is identical in general structure with that of *Ovis vignei*. The G-band pattern of the biarmed chromosomes was indistinguishable from the largest biarmed autosomal pair (pair I) of *O. musimon*, *O. orientalis*, *O. canadensis*, and *O. musimon*  $\times$  *O. canadensis*  $F_1$  and  $F_2$  hybrids. Among the acrocentric chromosomes of *Ammotragus* were pairs with G-band patterns resembling those seen in biarmed pairs II and III of  $2N = 54$  wild and domestic sheep. Buckland and Evans (1978a) reported that, with the exception of a single Robertsonian translocation, the G-band pattern is the same as the goat karyotype, and there is virtually no demonstrable centromeric heterochromatin (Buckland and Evans, 1978b). Karyotypes were published by Heck et al. (1968), Schmitt and Ulbrich (1968), and Nadler et al. (1974).

The  $F_1$  hybrid from a natural breeding of a male with a female domestic goat, *Capra hircus* ( $2N = 60$ ), was a  $2N = 59$  female with a karyotype containing one unpaired metacentric, two unpaired and 27 paired acrocentric autosomes, and two large acrocentric X's (Bunch et al., 1977). The phenotype incorporated features from both species.

Several studies intended to clarify the phylogenetic relationship of *Ammotragus* to the other Caprini may have only intensified the controversy. Serum protein analyses by Schmitt (1963) and immunoglobulin cross-reactivity studies by Curtain and Fudenberg (1973) indicated a close relationship between *Ammotragus* and *Ovis*. The amino acid sequence of various hemoglobin chains examined by Manwell and Baker (1975) showed that *Ammotragus* hemoglobin was more nearly similar to that from the domestic goat, *Capra hircus*, than from the domestic sheep, *Ovis aries*, but also exhibited some unique characteristics. But an immunological technique adapted for computer analysis by Hight and Nadler (1976) demonstrated a closer relationship between *Ovis* and *Capra* than between either of these and *Ammotragus*.

**REMARKS.** *Ammotragus* may have shared the epithet, *Ophion*, with *Ovis ammon* in ancient accounts by Pliny and others (Cuvier, 1827:359). The species was called *Tragelaphus* by Caius in 1561 (Cuvier, 1827:319; Scortecci, 1957:492), and *Lerwee* or *Fish-tall* by Shaw (1738). *Fish-tall* may derive from *fish*, bristles or mane, and *tall*, long, hence long mane; or from *vehsh*, a desert, and *tall*, wanderer (Cuvier, 1827:320). Common native appellations include *aoudad* (Mauretania), *kebsh* (Egypt), and *beddan* (males only) or *tedal/teytal* (Nubia) (Blyth, 1839:76).

A succession of generic or subgeneric reassessments, which do not properly belong in the synonymy, may have contributed to confusion about the taxonomy of this species. This nomenclature is included here for the convenience of interested persons.

*Ovis (Ammotragus) tragelaphus* Blyth, 1840:13.

*Ammotragus tragelaphus* Gray, 1850:40.

*Musimon tragelaphus* Gervais, 1855:192.

*Aries tragelaphus* Lataste, 1887:35.

*Ovis (Ammotragus) lervia* Lydekker, 1898:226.

*Ovis lervia* Anderson and de Winton, 1902:334.

Because Barbary sheep sometimes sire live hybrid offspring when bred to goats, and appear to be closer to goats in gross anatomical characteristics, Ansell (1971), Van Gelder (1977), and Corbet (1978) have placed Barbary sheep in the genus *Capra*. We demur from this assignment since the phylogenetic relationships are ambiguous, as noted in the section on genetics.

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#### LITERATURE CITED

Afzelius, A. 1815. De Antilopis in genere et speciation Guineensibus. *Nova Acta Regiae Soc. Sci. Upsala*, 7:195-270.

Allen, G. M. 1939. A checklist of African mammals. *Bull. Mus. Comp. Zool.*, 83:548.

Allen, R. W. 1960. Diseases and parasites of Barbary and big-horn sheep in the Southwest. *Trans. Desert Bighorn Council*, 4:17-22.

Allen, R. W., W. W. Becklund, and R. E. Gilmore. 1956. Parasites of the Barbary sheep. *J. Parasitol.*, 42(4 sect. 2):19.

Allen, R. W., G. A. Schad, and K. S. Samson. 1958. Experimental cross-transmission of two strains of *Haemonchus* from wild ruminants to domestic sheep, with observations on their pathogenicity as compared with *Haemonchus* from domestic sheep. *J. Parasitol.*, 44(4 sect. 2): 26.

Anderson, J., and W. E. de Winton. 1902. *Mammalia. Zoology of Egypt*, vol. 2, London, xvii + 374 pp.

Ansell, W. F. H. 1971. Order Artiodactyla. Pp. 1-84, in *The mammals of Africa, an identification manual* (J. Meester and H. W. Setzer, eds.). Smithsonian Institution Press, Washington.

Arambourg, C., M. Boule, H. Vallois, and R. Verneau. 1934. Les grottes paléolithiques des Beni Segoual, Algérie. Institut de paléontologie humaine, Paris. *Archives, Mém.*, 13:1-242.

Barrett, R. H. 1967. Some comparisons between the Barbary sheep and the desert bighorn. *Trans. Desert Bighorn Council*, 11:16-26.

Bate, D. M. A. 1955. Vertebrate faunas of Quaternary deposits in Cyrenaica. Pp. 274-291, in *Prehistory and Pleistocene geology in Cyrenaican Libya* (C. B. M. McBurney and R. W. Hey, eds.). Cambridge Univ. Press, Cambridge, 308 pp.

Bigourdan, J., and R. Prunier. 1937. Les mammifères sauvages de l'ouest Africain et leur milieu. *Encycl. Biol.*, Paris, 23:182-184.

Blunt, F. 1963. Barbary sheep . . . not in Wyoming, we hope. *Wyoming Wildl.*, 27(6):28-31.

Blyth, E. 1839. An amended list of the species of the genus *Ovis*. *Proc. Zool. Soc. London*, 7:62-79.

— 1840. A summary monograph of the species of the genus *Ovis*. *Proc. Zool. Soc. London*, 8:12-13.

Boever, W. J. 1976. Johne's disease in aoudads and mouflon. *J. Zoo Anim. Med.*, 7(1):19-23.

Bourdelle, E. 1924. Quelques caractères anatomiques du Mouflon à manchettes. *Revue Hist. Nat. Appl.*, Paris, 5:129-140.

Brack, M. 1966. Mycoplasmosis bei jungen Mähnenpringern (*Ammotragus lervia*). *Berl. Münch. Tierärztl. Wschr.*, Berlin, 79:169-172.

Brady, P. S., and D. E. Ullrey. 1975. Protein requirement of weaned aoudads (*Ammotragus lervia*). *J. Zool. Anim. Med.*, 6(4):24-28.

Brentjes, B. 1968. Bastardierung von Mähnen schaf, *Ammotragus lervia* (Pallas, 1777) und Hausziege, *Capra hircus* (Linné, 1758), in Altägypten? *Saeugetierk. Mitt.*, 16:153-157.

Brouin, G. 1950. Contribution à l'étude de l'Air. Notes sur les ongulés du cercle d'Agadez et leur chasse. *Mém. Soc. Hist. Nat. Afr. N.*, Alger, 10:425-455.

Brown, C. E. 1936. Rearing wild animals in captivity, and gestation periods. *J. Mamm.*, 17:10-13.

Buckland, R. A., and H. J. Evans. 1978a. Cytogenetic aspects of phylogeny in the Bovidae. I. G-banding. *Cytogenet. Cell Genet.*, 21:42-63.

— 1978b. Cytogenetic aspects of phylogeny in the Bovidae. II. C-banding. *Cytogenet. Cell Genet.*, 21:64-71.

Bunch, T. D., A. Rogers, and W. C. Foote. 1977. G-band and transferrin analysis of aoudad-goat hybrids. *J. Hered.*, 68:210-212.

Clark, J. L. 1964. The great arc of the wild sheep. *Univ. Oklahoma Press*, Norman, 247 pp.

Corbet, G. B. 1978. The mammals of the Palaearctic region: a taxonomic review. *British Museum (Natural History)* and Cornell Univ. Press, London and Ithaca, 314 pp.

Curtain, C. C., and H. H. Fudenberg. 1973. Evolution of the immunoglobulin antigens in the Ruminantia. *Biochem. Genet.*, 8:301-308.

Cuvier, G. 1827. The animal kingdom, arranged in conformity with its organization . . . (C. Griffith et al., compilers) G. B. Whittaker, London, 5:1-391.

Davis, W. B. 1974. The mammals of Texas. *Texas Parks and Wildl. Dept., Austin, Bull.*, 41:1-294.

DeArment, R. 1971. Reaction and adaptability of introduced aoudad sheep. Final Report. Div. of Fed. Aid in Wildlife Restoration Proj. No. W-45-R-21, Texas Parks and Wildl. Dept., Austin, 20 pp.

Edmond-Blanc, F. 1957. Le grand livre de la faune Africaine et de sa chasse, vol. 1. Rene Kister, Geneva.

Ernø, H., et al. 1972. The identification of an organism isolated from maned sheep as *Mycoplasma mycooides* subsp. *mycooides*. *Acta Vet. Scand.*, Copenhagen, 13:263-265.

Evans, P. K. 1967. The aoudad sheep, an exotic introduced in the Palo Duro Canyon of Texas. *Proc. Southeastern Assoc. Game Fish Comm.*, 21:183-188.

Flower, S. 1932. Notes on the recent mammals of Egypt, with a list of the species reported from that kingdom. *J. Zool.*, London, 2:369-450.

Geist, V. 1966. On the behavior and evolution of American mountain sheep. Ph.D. dissert., Univ. British Columbia, Vancouver, 251 pp.

Geoffroy Saint-Hilaire, I. 1827. Le Mouflon à manchettes. Pp. 264-268, in *Dictionnaire classique d'histoire naturelle*, 2nd ed. (J. B. G. M. Bory de Saint-Vincent, ed.). Mus. Nat. Hist., Paris.

Gervais, P. 1852. *Zoologique et paleontologie Françaises* . . . Arthur Bertrand, Paris.

— 1855. *Histoire naturelle des mammifères* . . . L. Curmer, Paris, 2:1-344.

Gray, A. P. 1954. Mammalian hybrids; a checklist with bibliography. *Commonwealth Agric. Bur. Farnham Royal*, Bucks, England, 144 pp.

Gray, G. C., D. B. Pence, and C. D. Simpson. 1978. Helminths of sympatric Barbary sheep and mule deer in the Texas Panhandle. *Proc. Helminthol. Soc. Washington*, 45:139-141.

Gray, J. E. 1850. Gleanings from the menagerie and aviary at Knowsley Hall. *Hoofed quadrupeds*. Publ. by the author, Knowsley, England, 2:1-76 + 4.

Haas, G. 1959. Untersuchungen über angeborene Verhaltensweisen bei Mähnenpringern (*Ammotragus lervia* Pallas). *Z. Tierpsychol.*, 16:218-242.

Hamdy, A. R., and T. Schmidt. 1972. Aggressive and sexual behavior among the Barbary sheep or aoudads (*Ammotragus lervia*) of the herd of the Egyptian Zoo in Cairo. *Bull. Fac. Sci., Cairo Univ.*, 45:277-283.

Hampy, D. B. 1978. Home range and seasonal movement of Barbary sheep in the Palo Duro Canyon. Unpubl. M.S. thesis, Texas Tech Univ., Lubbock, 83 pp.

Hampy, D. B., D. B. Pence, and C. D. Simpson. 1979. Serological

logical studies on sympatric Barbary sheep and mule deer from Palo Duro Canyon, Texas. *J. Wildl. Dis.*, 14:443-446.

Harper, F. 1940. The nomenclature and type localities of certain Old World mammals. *J. Mamm.*, 21:322-332.

Heck, H., D. Wurster, and K. Benirschke. 1968. Chromosome study of members of the subfamilies Caprinae and Bovinae, family Bovidae; the musk ox, ibex, aoudad, Congo buffalo, and gaur. *Z. Säugetierk.*, 33:172-179.

Heinemann, D. 1972. Barbary sheep, tahr, and blue sheep. Pp. 492-496, in Grzimek's animal life encyclopedia (B. Grzimek, ed.), Von Nostrand Reinhold, New York, 13(4):1-566.

Heughlin, T. 1861. Forschungen über die Fauna des Rothen Meers und der Somáli-Küste. Pp. 11-32, in Petermann's Mittheilungen aus Justus Perthes' geographischer anstalt (A. Petermann, ed.), Justus Perthes, Gotha, 482 pp.

Hight, M. E., and C. F. Nadler. 1976. Relationships between wild sheep and goats and the aoudad (Caprini) studied by immuno-diffusion. *Comp. Biochem. Physiol.*, 54B:265-269.

Huisman, T. H. J. 1974. Structural aspects of fetal and adult hemoglobins from nonanemic ruminants. *Ann. New York Acad. Sci.*, 241:392-410.

Huisman, T. H. J., et al. 1968. Studies of haemoglobin types in Barbary sheep (*Ammotragus lervia*). *Biochem. J.*, 107:745-751.

Huisman, T. H. J., and A. Miller. 1972. Hemoglobin types in Barbary sheep (*Ammotragus lervia* Pallas, 1777); absence of a  $\beta^{(cns)}$  animal during severe anemia. *Proc. Soc. Exp. Biol. Med.*, 140:815-819.

Huisman, T. H. J., G. van Vliet, and T. Sebens. 1958. Haemoglobin types in different species of sheep. *Nature*, 182:172-174.

Joleaud, L. 1927. Etudes de géographie zoologique sur la Berbérie: Le mouflon à manchettes. *C. R. Somm. Séanc. Soc. Biogéogr.*, Paris, 27:43-45.

Katz, I. 1949. Behavioral interactions in a herd of Barbary sheep (*Ammotragus lervia*). *Zoologica*, 34(3):9-18.

Kéler, S. 1942. Ein Beitrag zur Kenntnis der Mallophagen. *Arb. Morphol. Taxon. Entomol. Berlin-Dahlem*, 9:69-85.

Lataste, F. 1887. Catalogue critique des mammifères apélagiques sauvages de la Tunisie. Imprimerie Nationale, Paris, 42 pp.

Lepri, G. 1930. Sopra una nuova sottospecie del genere *Ammotragus*. *Atti Pont. Acad. Sci. Nuovi Lincei*, Roma, 83:271.

Lhote, H. 1957. Le grand livre de la faune Africaine et de sa chasse, vol. 2. Rene Kister, Geneva.

Lobanov, N. V., and V. D. Treus. 1971. [Reproduction of *Ammotragus lervia* at the Askaniya-Nova Zoo.] *Vest. Zool.*, 5(3):23-26. [In Russian with English summary]

Lydekker, R. 1898. Wild oxen, sheep, and goats of all lands, living and extinct. Rowland Ward, London, 318 pp.

— 1912. The sheep and its cousins. Ballantyne, Hanson & Co., Edinburgh, 315 pp.

— 1913. Catalogue of the ungulate mammals in the British Museum (Natural History). British Museum, London, 1:1-249.

Malbrant, R. 1952. Faune du Centre Africain Français (mammifères et oiseaux). Encycl. Biol., Paris, 2nd ed., 15:59-61.

Manwell, C., and C. M. A. Baker. 1975. *Ammotragus lervia*: progenitor of the domesticated sheep or specialized offshoot of caprine evolution? *Experientia*, 31:1370-1371.

— 1977. *Ammotragus lervia*: Barbary sheep or Barbary goat? *Comp. Biochem. Physiol.*, 58B:267-271.

McBurney, C. B. M. 1967. The Haua Fteah (Cyrenica) and the Stone Age of the south-east Mediterranean. Cambridge Univ. Press, Cambridge, 387 pp.

McClellan, J. 1955. Preliminary report on the Barbary sheep of New Mexico. *Proc. Western Assoc. State Game Fish Comm.*, 35:167-176.

Meyer, H. 1967. Beobachtungen über Hämoglobine und Blutkaliumwerte bei Mähnenschafen (*Ammotragus lervia*) und Tahren (*Hemitragus jemlahicus*). *Z. Säugetierk.*, 32:178-180.

Nadler, C. F., R. S. Hoffmann, and A. Woolf. 1974. G-band patterns, chromosomal homologies, and evolutionary relationships among wild sheep, goats, and aoudads (Mammalia, Artiodactyla). *Experientia*, 30:744-746.

Nicholaisen, J. 1963. Ecology and culture of the pastoral Tuareg, with particular reference to the Tuareg of Ahaggar and Ayr. National Museum, Copenhagen, Etnografisk Raekke, 9:1-548.

Nouvel, J., G. Chauvier, and L. Strazielle. 1969. Effects de quelques Tranquillisants et Anesthésiques sur les Animaux sauvages. *Zool. Garten*, 37:114-116.

Ogren, H. A. 1962. The Barbary sheep, *Ammotragus lervia* (Pallas), of the Canadian River Gorge, New Mexico. Unpubl. Ph.D. dissert., Univ. Southern California, Los Angeles, 256 pp.

— 1965. Barbary sheep. *New Mexico Dept. Fish & Game*, Santa Fe, Bull. 13:1-117.

Osborn, H. F. 1910. The age of mammals in Europe, Asia and North America. MacMillan, New York, 635 pp.

Pallas, P. S. 1777. *Antilope lervia*. *Spicilegia Zool.*, 12:12.

Panouse, J. B. 1957. Les mammifères du Maroc: primates, carnivores, pinnipèdes, artiodactyles. *Trav. Inst. Scient. Chérif., Tanger*, Ser. Zool., 5:169-173.

Payne, S. 1968. The origins of domestic sheep and goats: a reconsideration in the light of the fossil evidence. *Proc. Prehist. Soc.*, 34:368-384.

Petzsch, H. 1957. Lebender ♀—Bastard aus *Ammotragus lervia* Pall.—♂ × *Capra hircus* L.—♀ im Berg-Zoo Halles-S. geboren. *Zool. Anzeiger*, 159:285-290.

Ramsey, C. W., and M. J. Anderegg. 1972. Food habits of an aoudad sheep, *Ammotragus lervia* (Bovidae), in the Edwards Plateau of Texas. *Southwestern Nat.*, 16:267-280.

Rode, P. 1943. Faune de l'Empire Français II. Mammifères ongules de l'Afrique Noire, Part I: Famille des Bovidés, genus *Ammotragus* Blyth, 1840. *Faune Tropicale*, 2:31-34.

Rothschild, W. 1913. On *Ovis lervia* Pallas and its subspecies. *Novit. Zool.*, 20:459.

— 1921. Captain Angus Buchanan's Air expedition. III. Ungulate mammals. *Novit. Zool.*, 28:75.

Samson, K. S., R. W. Allen, and G. A. Schad. 1964. Comparative pathogenicity in *Ovis aries* of homologous and heterologous strains of *Haemonchus* (Nematoda: Trichostrongylidae) from domestic and wild sheep. *J. Parasitol.*, 50:421-426.

Schaffer, W. M. 1968. Intraspecific combat and the evolution of the Caprini. *Evolution*, 22:817-825.

Schaffer, W. M., and C. A. Reed. 1972. The co-evolution of social behavior and cranial morphology in sheep and goats (Bovidae, Caprini). *Fieldiana Zool.*, Chicago, 61:1-88.

Schmitt, J. 1963. *Ammotragus lervia* Pallas, Mähnenschaf oder Mahnenziege? *Z. Säugetierk.*, 28:7-12.

Schmitt, J., and F. Ulbrich. 1968. Die Chromosomen verschiedener Caprini Simpson, 1945. *Z. Säugetierk.*, 33:180-186.

Schreiner, C., III. 1968. Uses of exotic animals in a commercial hunting program. Pp. 13-16, in Caesar Kleberg Research Program in Wildlife Ecology, Symposium on Introduction of exotic animals: ecological and socioeconomic considerations. Coll. Agric., Texas A&M Univ., College Station, 30 pp.

Scortecci, G. 1957. Animali. Vol. 1, Mammiferi. Edizioni Labor, Milano.

Serres, M. de. 1848. Fossil bones. Cavernes, Paris.

Shae, T. 1738. Travels, or observations relating to several parts of Barbary and the Levant. Printed at the Theatre, Oxford, 442 pp.

Simpson, C. D., et al. 1978. The Barbary sheep: a threat to desert bighorn survival. *Trans. Desert Bighorn Council*, 22:26-31.

Steklenev, E. P. 1972. [Morphological characterization of sperm from representatives of the subfamily Caprinae in relation to their hybridization.] *Sel'-khoz. Biol.*, 7:500-506.

Strasson, O. (ed.). 1916. Brehm's Tierleben. Allgemeine kunde des tierreichs. Säugetiere-Vierte Band. Bibliogr. Inst., Leipzig und Wien.

Teer, J. G. 1975. Commercial uses of game animals on rangelands of Texas. *J. Anim. Sci.*, 40:1000-1008.

Thomas, O. 1902. On the mammals collected during the Whittaker Expedition to Tripoli. *Proc. Zool. Soc. London*, 1902(2): 1-13.

Trainer, D. O., and M. M. Jochim. 1969. Serologic evidence of bluetongue in wild ruminants of North America. *Amer. J. Vet. Res.*, 30:2007-2011.

Tumbleston, M. E., C. C. Middleton, and J. D. Wallach. 1970. Serum biochemical and hematologic parameters of adult aoudads (*Ammotragus lervia*) in captivity. *Lab. Anim. Care*, 20:242-245.

Van Gelder, R. G. 1977. Mammalian hybrids and generic limits. *Amer. Mus. Novitates*, 2635:1-25.

Vaufrey, R. 1955. Préhistoire de l'Afrique. I. Le Maghreb. *Publ. Inst. Hautes Etudes Tunis*, Masson, Paris, 4:1-435.

Wallach, J. D., and C. C. Middleton. 1970. Naturally occur-

ring atherosclerosis in aoudads (*Ammotragus lervia* (Pallas)). *Acta Zool. Pathol. Antverpiensia*, 50:45-54.

Werneck, F. L. 1950. *Os Malófagos de Mamíferos. Parte II: Ischnocera (continuação de Trichodectidae) e Rhyncophthirina*. Ed. Inst. Oswaldo Cruz, Rio de Janeiro, 207 pp.

Zammarano, V. T. 1930. *Fauna e caccia*. Minist. Colonie, Rome, pp. 24-26.

Zuckerman, S. 1953. The breeding seasons of mammals in captivity. *Proc. Zool. Soc. London*, 122:827-949.

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